

consistent with Elejalde syndrome (neuroectodermal melanolyosomal disorder). *J Neurol* 247:570–572  
 Seabra MC, Mules EH, Hume AN (2002) Rab GTPases, intracellular traffic and disease. *Trends Mol Med* 8:23–30

Address for correspondence and reprints: Dr. Geneviève de Saint Basile, INSERM U429, Hôpital Necker-Enfants Malades, 169 rue de Sevres, 75015 Paris, France. E-mail: sbasile@necker.fr

© 2002 by The American Society of Human Genetics. All rights reserved. 0002-9297/2002/7105-0026\$15.00

Ménasché G, Fischer A, de Sainte Basile G (2002) Griscelli syndrome type 1 and 2. *Am J Hum Genet* 71:1237–1238 (in this issue)

Address for correspondence and reprints: Dr. Marjan Huizing, 10 Center Drive, MSC 1851, Building 10, Room 10C-103, National Human Genome Research Institute, National Institutes of Health, Bethesda, MD 20892-1851. E-mail: mhuizing@mail.nih.gov

© 2002 by The American Society of Human Genetics. All rights reserved. 0002-9297/2002/7105-0027\$15.00

*Am. J. Hum. Genet.* 71:1238, 2002

### Reply to Ménasché et al.

*To the Editor:*

It is gratifying to learn that Ménasché et al. (2002 [in this issue]) agree with our analysis of the phenotypic differences between patients with *RAB27A* mutations and those with *MYO5A* mutations. We leave it to *Journal* readers to decide if previous publications have “unequivocally established” these points. We do apologize for the error in table 1, which we recognized and have corrected in an erratum.

Perhaps we could make two additional points. First, Gaucher disease types I, II, and III represent examples of defects in a single gene resulting in different phenotypes, whereas Griscelli/Elejalde syndromes represent examples of defects in two different genes resulting in phenotypes with some similarities. Second, we wonder what nomenclature should be employed for these two disorders. Ménasché et al. continue to use Griscelli syndromes types 1 and 2. However, Griscelli’s original cases exhibited immune deficiency (Griscelli et al. 1978), whereas Elejalde first recognized a distinct, neurologically based disorder (Elejalde et al. 1979). Perhaps Dr. Elejalde should be credited for the accuracy of his ascertainment.

MARJAN HUIZING,<sup>1</sup> Y. ANIKSTER,<sup>2</sup> AND W. A. GAHL<sup>1</sup>  
<sup>1</sup>*Section on Human Biochemical Genetics, Medical Genetics Branch, National Human Genome Research Institute, National Institutes of Health, Bethesda, MD; and* <sup>2</sup>*Metabolic Unit, Sheba Medical Center, Tel Hashomer, Israel*

### References

- Elejalde BR, Holguin J, Valencia A, Gilbert EF, Molina J, Marin G, Arango LA (1979) Mutations affecting pigmentation in man. I. Neuroectodermal melanolyosomal disease. *Am J Med Genet* 3:65–80  
 Griscelli C, Durandy A, Guy-Grand D, Daguillard F, Herzog C, Prunieras M. (1978) A syndrome associating partial albinism and immunodeficiency. *Am J Med* 65:691–702

*Am. J. Hum. Genet.* 71:1238–1239, 2002

### Family-Based Association Tests Incorporating Parental Genotypes

*To the Editor:*

The report “Parental Genotypes in the Risk of a Complex Disease” (Labuda et al. 2002) gives several interesting examples of how parental genotypes can contribute to children’s disease risk—for example, through maternal effects during pregnancy or paternal effects during spermatogenesis. The authors note that if disease risk depends on parents’ genotypes but not their child’s genotype, then the distribution of genotypes in cases will not differ from the Mendelian expectation given their parents’ genotypes. Hence, the traditional transmission disequilibrium test (TDT) using case-parent trio data will (correctly) not detect any association between individuals’ genotypes and disease. The authors present an example in which the TDT provides no evidence of an association between a variant allele and disease (in fact, the point estimate for the odds ratio is 1.0), whereas a comparison of case subjects’ genotypes to those of population control subjects does provide evidence of association (estimated odds ratio = 3.4). The authors then compare maternal and paternal genotypes to control subjects’ genotypes and find evidence that the prevalence of the variant allele is higher in parents of case subjects than in population control subjects.

However, there are other analytic options in this case—namely, flexible statistical methods for case-parent trios which can test for parental-genotype effects. These have the advantage of being robust to population-stratification bias and, in some situations, are even more powerful for testing for parental-genotype effects than case-control studies (Starr et al. 2002).

The log-linear model developed by Weinberg et al. and Wilcox et al. can test for parental-genotype and parent-of-origin effects after adjusting for possible case-genotype effects (Weinberg et al. 1998; Wilcox et al. 1998). In principle, this model can also test parental-genotype × case-genotype interactions—which could be relevant;

for example, in the study of complications during pregnancy such as preeclampsia (Kilpatrick 1999).

The log-linear model is equivalent to a conditional logistic analysis comparing the case to appropriately defined “pseudo-sibling controls” (Kraft 2002). If the gene under study is assumed not to play a direct role in an individual’s risk of disease (or to be linked to any other such gene), then to test for an indirect role of maternal genotype (say) each case subject should be compared to a pseudo-sibling control subject whose mother has the genotype of the case subject’s father. That is, if the genotypes of the mother and father are  $G_m$  and  $G_f$ , respectively, then the conditional logistic likelihood for the family is

$$\frac{e^{\beta Z(G_m)}}{e^{\beta Z(G_m)} + e^{\beta Z(G_f)}} ,$$

where  $Z(\cdot)$  is some dominance coding.

This approach (reasonably) assumes that, given the set of parental marker genotypes  $\{G_1, G_2\}$ , it is equally likely that  $G_m = G_1$  or  $G_2$ . In other words, “the frequency of heterozygous mothers married to homozygous variant fathers is the same as the frequency of heterozygous fathers married to homozygous variant mothers, and so on” (Wilcox et al. 1998). Furthermore, since this likelihood permutes the genotypes of “the parent contributing to disease risk” and the “the parent not contributing to disease risk,” it cannot estimate joint effects of both parents’ genotypes. However, for many diseases, only the mother (father) will plausibly contribute to a child’s disease risk.

Although the case-parent trio analysis conditions on the parents’ genotypes and hence is robust to population stratification bias, the analysis comparing parental genotypes to population-based controls is not (although Labuda et al. [2002] argue that this may not be an issue for the particular data they analyze in their report). Furthermore, even when there is no population stratification, the latter analysis is something of an “apples and oranges” comparison, as the exposure of interest is not the control subject’s genotype, but his or her parent’s genotype. The control’s genotype serves as a surrogate for his or her parent’s. In a simulation study with 175 unmatched case and control subjects (1,000 replicates), we found that the odds ratio comparing case subjects’ maternal genotypes to control genotypes underestimated the odds ratio associated with each variant maternal allele by 11% (variant allele frequency 0.25; baseline probability of disease 14%; odds ratio per variant maternal allele 2). Of course, the data Labuda et al. (2002) analyzed did not contain parental genotype information for the controls. But if one were to design a case-population control study to detect the effects of maternal

(paternal) genotypes, then one should plan to collect information on controls’ maternal (paternal) genotypes.

Finally, figure 1a is misleading in that case subjects’ parents are not representative of population controls if individuals’ genotypes are associated with disease or there is population stratification.

PETER KRAFT AND MELISSA WILSON

*Department of Preventive Medicine  
Keck School of Medicine  
University of Southern California  
Los Angeles*

## References

- Kilpatrick D (1999) Influence of human leukocyte antigen and tumor necrosis factor genes on the development of preeclampsia. *Hum Reprod Update* 5:94–102
- Kraft P (2002) Statistical methods in family-based gene-association studies. PhD dissertation, University of Southern California, Los Angeles
- Labuda D, Krajinovic M, Sabbagh A, Infante Rivard C, Sinnett D (2002) Parental genotypes in the risk of a complex disease. *Am J Hum Genet* 71:193–197
- Starr J, Hsu L, Schwartz SM (2002) Maternal genetics as risk factors for disease in offspring: statistical power of the log-linear approach to case-parent triads vs. a case-control design. *Am J Epidemiol* 155:S50
- Weinberg C, Wilcox A, Lie RT (1998) A log linear approach to case-parent data: assessing effects of disease genes that act directly or through maternal effects, and may be subject to parental imprinting. *Am J Hum Genet* 62:969–978
- Wilcox A, Weinberg C, Lie RT (1998) Distinguishing the effects of maternal and offspring genes through studies of “case parent triads.” *Am J Epidemiol* 148:893–901

Address for correspondence and reprints: Dr. Peter Kraft, University of Southern California, 1540 Alcazar Street, CHP 218 MC 9010, Los Angeles, CA 90089-9010. E-mail: pkraft@usc.edu

© 2002 by The American Society of Human Genetics. All rights reserved.  
0002-9297/2002/7105-0028\$15.00

*Am. J. Hum. Genet.* 71:1239–1240, 2002

## Regarding “Parental Genotypes in the Risk of a Complex Disease”

*To the Editor:*

Labuda et al. (2002) have proposed that parental genotypes might play a role in the causation of complex diseases. They seem unaware that this idea has been considered by others (e.g., Lande et al. 1989) and that methods have been developed to test for parentally mediated genetic effects, both for a dichotomous phenotype (Mitchell 1997; Weinberg et al. 1998; Wilcox et al.